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Infinium LCG Quad Assay
Lab Tracking Form

1 Make and Read Quant (Optional/LIMS)

Date/Time: _____
Operator: _____ Robot: _____

WG#-DNA Plate: QNT Plate:
(1) _____ _____
(2) _____ _____
(3) _____ _____

Standard DNA Plate: Standard QNT Plate:
_____ _____

2 Make AMP5

Date/Time: _____
Operator: _____ Robot: _____
Plate Positions on Robot Bed: _____
Batch #: _____
Number of Samples: _____
Hyb oven (37°C, 20-24 h): Start _____ Stop _____
WG#-DNA Plate: _____
0.1N NaOH Reagent: _____
AMP5 Plate: _____

RPM Reagent: (Col. 1) _____
(Col. 3) _____
(Col. 5) _____
(Col. 7) _____
(Col. 9) _____
(Col. 11) _____
AMM Reagent: (Col. 1) _____
(Col. 3) _____
(Col. 5) _____
(Col. 7) _____
(Col. 9) _____
(Col. 11) _____

Column 1
Well: Sample
A01: _____
B01: _____
C01: _____
D01: _____
E01: _____
F01: _____
G01: _____
H01: _____

Column 3
Well: Sample
A03: _____
B03: _____
C03: _____
D03: _____
E03: _____
F03: _____
G03: _____
H03: _____

Column 5
Well: Sample
A05: _____
B05: _____
C05: _____
D05: _____
E05: _____
F05: _____
G05: _____
H05: _____

Column 7
Well: Sample
A07: _____
B07: _____
C07: _____
D07: _____
E07: _____
F07: _____
G07: _____
H07: _____

Column 9
Well: Sample
A09: _____
B09: _____
C09: _____
D09: _____
E09: _____
F09: _____
G09: _____
H09: _____

Column 11
Well: Sample
A011: _____
B011: _____
C011: _____
D011: _____
E011: _____
F011: _____
G011: _____
H011: _____

Project: _____
Batch: _____
AMP5 Plate: _____
Image Date: _____

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3 Fragment AMP5

Date/Time: _____
Operator: _____ Robot: _____
Plate Positions on Robot Bed: _____
☐ Vortex at 1600 rpm
Heat block (37°C, 1 h): Start _____ Stop _____

FRG Reagent: _____
FRG Reagent: _____
FRG Reagent: _____

4 Precip AMP5

Date/Time: _____
Operator: _____ Robot: _____
Plate Positions on Robot Bed: _____
☐ Vortex at 1600 rpm
Heat block (37°C, 5 m): _____
Incubate (4°C, 30 m): Start: _____ Stop: _____
☐ Centrifuge AMP5 plate to 3000 xg (4°C, 20 m)
Air dry (22°C, 1 h): Start: _____ Stop: _____

2-propanol Lot #: _____
2-propanol Date Opened: _____
PA1 Reagent: _____
PA1 Reagent: _____
PA1 Reagent: _____

5 Resuspend AMP5

Date/Time: _____
Operator: _____ Robot: _____
Plate Positions on Robot Bed: _____
Hyb oven (48°C, 1 h): Start _____ Stop _____

RA1 Reagent: _____

6 Hyb Multi-Use BeadChip

Date/Time: _____
Operator: _____ Robot: _____
Heat block (95°C, 20 m): Start: _____ Stop: _____
Benchtop incubation (RT, 30 m): Start: _____ Stop: _____
☐ Centrifuge AMP5 plate to 280 xg
Hyb oven (48°C, 16-24 h): Start _____ Stop _____

PB2 Reagent: _____
_____
_____

Refer to page 4 for BeadChip loading instructions. Enter the BeadChip barcodes in the spaces provided.

7 Wash BeadChip

Date/Time: _____
Operator: _____

PB1 Reagent: _____
_____
_____

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8 XStain LCG BeadChip

Date/Time: _____
Operator: _____ Robot: _____
RA1 Reagent: _____
LX1 Reagent: (1-4) _____
(5-8) _____
(9-12) _____
(13-16) _____
(17-20) _____
(21-24) _____
LX2 Reagent: (1-4) _____
(5-8) _____
(9-12) _____
(13-16) _____
(17-20) _____
(21-24) _____
EML Reagent: (1-4) _____
(5-8) _____
(9-12) _____
(13-16) _____
(17-20) _____
(21-24) _____
95% formamide/1 mM EDTA Reagent: _____
XC3 Reagent: _____
SML Reagent: (1-4) _____
(5-8) _____
(9-12) _____
(13-16) _____
(17-20) _____
(21-24) _____
SML Temperature: _____
ATM Reagent: (1-4) _____
(5-8) _____
(9-12) _____
(13-16) _____
(17-20) _____
(21-24) _____
PB1 Reagent: _____
XC4 Reagent: _____

Record the chamber rack position for each BeadChip on page 5.

9 Image BeadChip

Record the barcode, Scanner ID, and the image date for each BeadChip on page 4.

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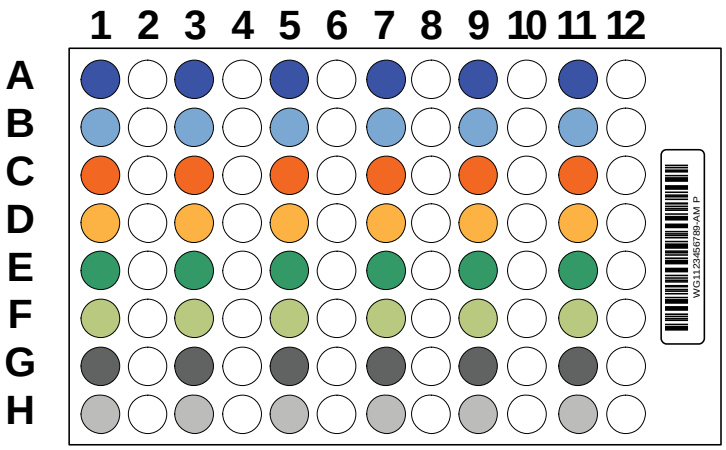
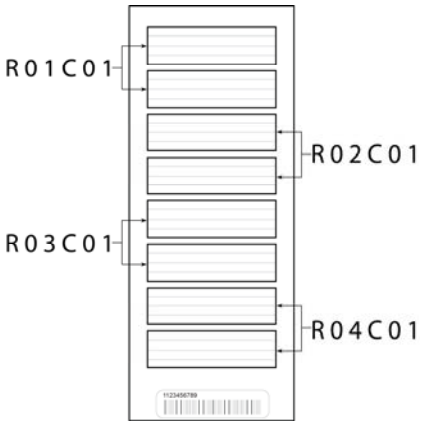
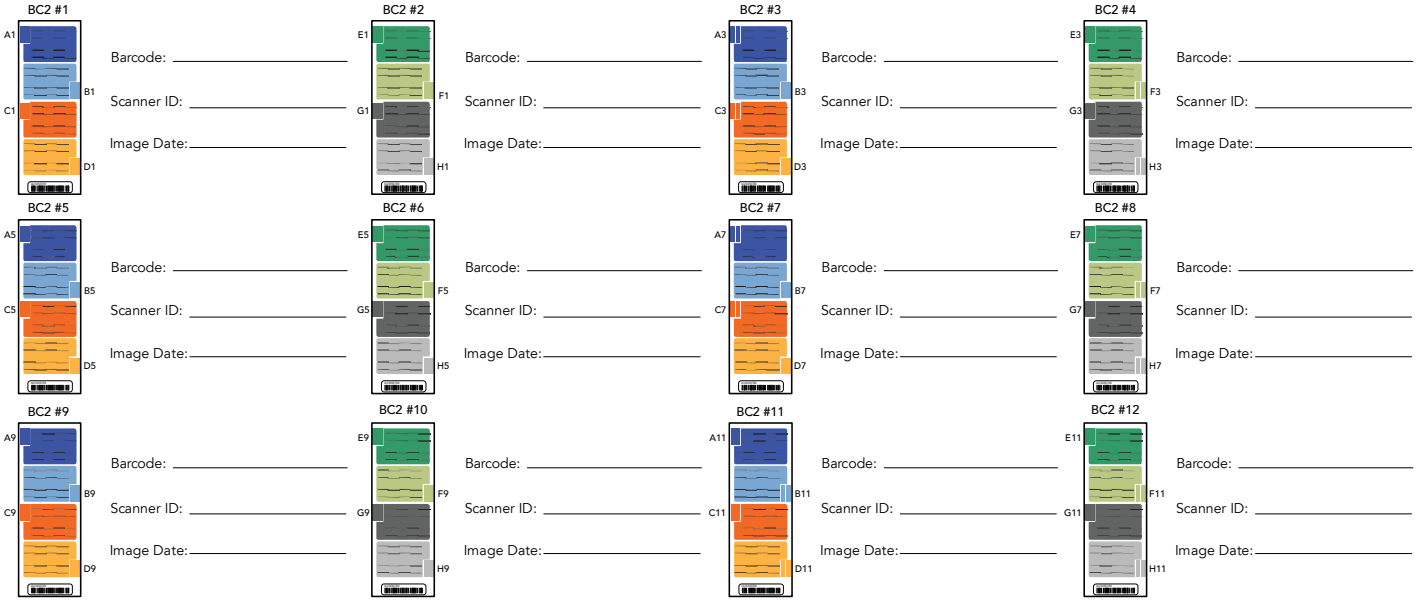
Track BeadChips

AMP Plate





Sample Section Naming Diagram



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Chamber Rack Position Chart

Use this chart to enter BeadChip IDs in the appropriate chamber rack position during the XStain LCG BeadChip step.

Row 1	Row 2	Row 3
1 _____	_____	17 _____
2 _____	10 _____	18 _____
3 _____	11 _____	19 _____
4 _____	12 _____	20 _____
5 _____	13 _____	21 _____
6 _____	14 _____	22 _____
7 _____	15 _____	23 _____
8 _____	16 _____	24 _____